

Cloning of oligonucleotides to S2 CRISPR vector

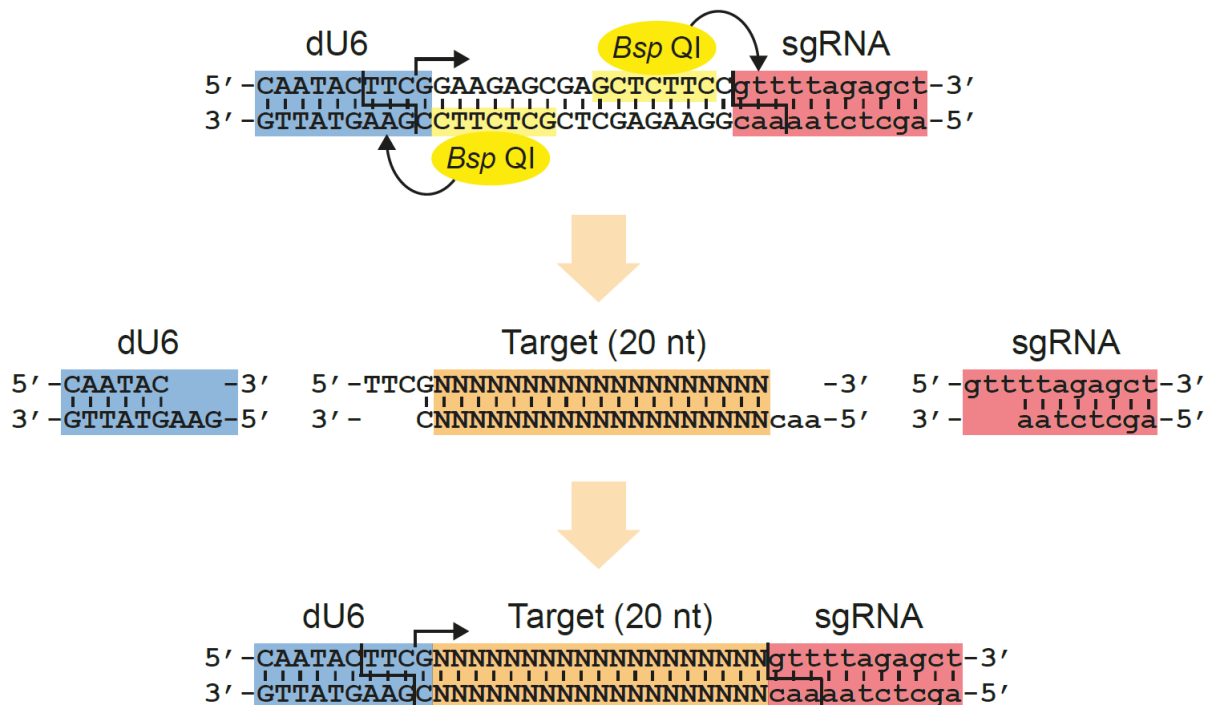
Andrew Bassett - 19th June 2013

1. Pick 23 nt sequence upstream of NGG in the genome
2. Remove NGG sequence to leave 20 nt target sequence
3. If target sequence does not begin with G, add G to the 5' end of it to allow transcription from the U6 promoter
4. For the forward oligonucleotide, add TTC to the 5' end of the target sequence
5. For the reverse oligonucleotide, reverse compliment the target sequence (including any additional G nucleotide), and add AAC to the 5' end
6. Synthesise two oligonucleotides

F = 5' - TTCGNNNNNNNNNNNNNNNNNNNNNN (N) -3'

R = 5' - AAC (N) NNNNNNNNNNNNNNNNNNNNNNC -3'

Schematic of cloning process



7. Anneal forward and reverse oligos as follows
8. Mix 10 μ l of 2x annealing buffer (20mM Tris, 2mM EDTA, 100mM NaCl, pH 8.0) with 5 μ l each of 100 uM forward and reverse oligos.
9. Anneal in PCR machine on the following program

98°C	1 min
98-88°C	5 s, decrease 0.1°C/cycle x99 cycles
88-78°C	10 s, decrease 0.1°C/cycle x99 cycles
78-68°C	10 s, decrease 0.1°C/cycle x99 cycles
68-58°C	10 s, decrease 0.1°C/cycle x99 cycles
58-48°C	10 s, decrease 0.1°C/cycle x99 cycles
48-38°C	10 s, decrease 0.1°C/cycle x99 cycles
38-18°C	1 s, decrease 0.2°C/cycle x99 cycles
18°C	Forever

10. Phosphorylate oligos in the following mixture

1 μ l	Annealed oligo
1 μ l	Roche T4 DNA ligase buffer (contains ATP)
1 μ l	T4 PNK
7 μ l	ddH ₂ O
<u>10 μl</u>	<u>Total</u>

11. Incubate for 30 min at 37°C

12. Dilute 10x in ddH₂O

13. Meanwhile, digest 2 μ g pAc-sgRNA-Cas9 vector for 1 h at 37°C

2 μ g	pAc-sgRNA-Cas9-Puro vector
5 μ l	10x NEBuffer 4
X μ l	ddH ₂ O
<u>2 μl</u>	<u>Bsp QI (NEB)</u>
50 μ l	Total

14. Add 1 μ l CIP (NEB) and incubate for 10 min at 37°C

15. PCR purify to 30 μ l EB

16. Ligate vector and oligo in the following mixture

1 μ l	Vector
2 μ l	10x diluted oligo
1 μ l	Roche T4 DNA ligase buffer
4 μ l	ddH ₂ O
<u>1 μl</u>	<u>T4 DNA ligase</u>
10 μ l	Total

17. Incubate at 18°C for 2 h, then transform 2 μ l to a 50 μ l aliquot of chemically competent DH5- α cells

a. Incubate 20 min on ice with plasmid

b. Heat shock for 30 s at 42°C

c. Cool for 2 min on ice

d. Add 150 μ l SOC and plate entire mixture to LB-carbenicillin plate (100 μ g/ml)

e. Incubate at 37°C overnight