From Methods Aakalu et al., (2001) Neuron 30(2): 489-502

Only steps relevant for pcDNA3.1-5′$_{myr}$dGFP3′

pcDNA3.1-5′dGFP3′: The CamKII-α 3′UTR sequence obtained from plasmid (Mayford et al., 1996) was PCR amplified (forward primer: 5′-ttatatttgcggccgcggtcgctaccattaccagtt-3′; reverse primer: 5′-ggcgctctctcgagtttaaatttgtagct-3′) and cloned into the NotI and XhoI sites of the pcDNA3.1 vector (Invitrogen). The resulting vector was then cleaved with BamHI and NotI for insertion of the destabilized EGFP ORF (from pd2EGFP, Clontech). The CamKII-α 5′UTR was released from a plasmid (obtained from J. Fallon) and inserted at the HindIII-BamHI sites, yielding pcDNA-5′dGFP3′.

pcDNA3.1-5′$_{myr}$dGFP3′: The d2EGFP ORF (from pd2EGFP, Clontech) was PCR amplified (forward primer: 5′-cgactctagagtgagcaagggcgaggagctg-3′; reverse primer: 5′-tctagagtcgcggccgcatctacaca-3′), digested, and inserted into the XbaI-NotI sites of pBSK. To generate the myristoylation signal, two oligos corresponding to the N-terminal 10 amino acids of p10 were annealed (myr1: 5′-gatccatgggcacggtgctgtccctgtctcccagct-3′; myr2: 5′-ctagagctgggagacagggacagcaccgtgcccatg-3′), digested, and inserted into the BamHI-XbaI sites of pBSK-d2EGFP. The $_{myr}$dGFP was subcloned into the BamHI-NotI sites of pcDNA3.1-5′dGFP3′.