

# A rapid sgRNA cloning protocol for CRISPRainbow (V20160427)

This protocol was adapted from Ma H *et al*, NBT, 2016. Hanhui Ma @ UMMS.

## A. Design sgRNA oligos

Fwd: ACCGNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN

Rev: AAACNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN

## B. Annealing oligos

### Reagents:

	Stock	ml
• Annealing Buffer		
100 mM Tris-HCl pH8.0	1 M	0.25
50 mM NaCl	5 M	0.25
1 mM EDTA	0.5 M	0.05
<u>ddH<sub>2</sub>O</u>		<u>24.5</u>
Total		25

- 100  $\mu$ M oligos

<u>Annealing:</u>	$\mu$ l
Annealing buffer	40
Fwd oligo (100 $\mu$ M)	5
<u>Rev oligo (100 <math>\mu</math>M)</u>	<u>5</u>
Total	50

- Incubate @ 95 °C for 3 min and slowly cool down to room temperature;
- Dilute 5  $\mu$ l of annealed oligos to 245  $\mu$ l water and final concentration is 200 nM (about 3ng/ $\mu$ l, ready for use).

## C. Cloning annealed oligos into vectors

### Reagents:

- pLH-sgRNA1 vectors\*
- Enzymes and Buffer in reaction
- Stbl3 competent cells
- LB-Ampicillin plates

<u>Reaction mix:</u>	$\mu$ l
pLH-sgRNA1 vector (100 ng/ $\mu$ l)	1
10XSmartCut Buffer (NEB)	1
10 mM ATP (NEB)	1
BbsI (NEB)	0.5
T7 DNA ligase (NEB)	0.3
ddH <sub>2</sub> O	5.2
<u>Annealed oligos (200 nM)</u>	<u>1</u>
	10

- Incubate @ 37 °C for 15 min;
- Transform 5  $\mu$ l of reaction mix into Stbl3 competent cells and spread on LB-Amp plates.

## D. Minipreps and Sequencing

### Reagents:

- QIAprep Spin Miniprep Kit
- Sequencing primer pLKO.1-Rseq:  
CTATTCTTTCCCCTGCACTGTACCC

### \* Notes of pLH-sgRNA1 vectors:

sgRNA1 carried a pair of mutations described as sgRNAplus in Supplementary Figure 1, Ma H *et al*, NBT, 2016.

All the sgRNA vector plasmids require CcdB Survival cells for growth.