

## Preparation of Penn State DNA ladders from pPSU1 and pPSU2 plasmids

### 1. analytical digest (each plasmid at $\sim 0.1 \mu\text{g}/\mu\text{l}$ ):

water	34 $\mu\text{l}$	water	33 $\mu\text{l}$
10x NEBuffer 3.1	5 $\mu\text{l}$	10x NEBuffer 3.1	5 $\mu\text{l}$
1 $\mu\text{g}/\mu\text{l}$ pPSU1	5 $\mu\text{l}$	1 $\mu\text{g}/\mu\text{l}$ pPSU1	5 $\mu\text{l}$
1 $\mu\text{g}/\mu\text{l}$ pPSU2	4 $\mu\text{l}$	1 $\mu\text{g}/\mu\text{l}$ pPSU2	5 $\mu\text{l}$
20 units/ $\mu\text{l}$ EcoRV	<u>2 <math>\mu\text{l}</math></u>	20 units/ $\mu\text{l}$ PstI	<u>2 <math>\mu\text{l}</math></u>
	50 $\mu\text{l}$		50 $\mu\text{l}$
water	17 $\mu\text{l}$		
10x NEBuffer 3.1	2.5 $\mu\text{l}$		
1 $\mu\text{g}/\mu\text{l}$ pPSU1	2.5 $\mu\text{l}$		
1 $\mu\text{g}/\mu\text{l}$ pPSU2	2 $\mu\text{l}$		
10 units/ $\mu\text{l}$ NcoI	<u>1 <math>\mu\text{l}</math></u>		
	25 $\mu\text{l}$		

digest at 37°C for 2 hours, check 1  $\mu\text{l}$  of digest on 1% agarose gel

### 2. preparative digest (each plasmid at $\sim 0.25 \mu\text{g}/\mu\text{l}$ ):

water	80 $\mu\text{l}$	water	70 $\mu\text{l}$
10x NEBuffer 3.1	20 $\mu\text{l}$	10x NEBuffer 3.1	20 $\mu\text{l}$
1 $\mu\text{g}/\mu\text{l}$ pPSU1	50 $\mu\text{l}$	1 $\mu\text{g}/\mu\text{l}$ pPSU1	50 $\mu\text{l}$
1 $\mu\text{g}/\mu\text{l}$ pPSU2	40 $\mu\text{l}$	1 $\mu\text{g}/\mu\text{l}$ pPSU2	50 $\mu\text{l}$
20 units/ $\mu\text{l}$ EcoRV	<u>10 <math>\mu\text{l}</math></u>	20 units/ $\mu\text{l}$ PstI	<u>10 <math>\mu\text{l}</math></u>
	200 $\mu\text{l}$		200 $\mu\text{l}$
water	40 $\mu\text{l}$		
10x NEBuffer 3.1	10 $\mu\text{l}$		
1 $\mu\text{g}/\mu\text{l}$ pPSU1	25 $\mu\text{l}$		
1 $\mu\text{g}/\mu\text{l}$ pPSU2	20 $\mu\text{l}$		
20 units/ $\mu\text{l}$ NcoI	<u>5 <math>\mu\text{l}</math></u>		
	100 $\mu\text{l}$		

digest at 37°C overnight, check 0.5  $\mu\text{l}$  of digest on 1% agarose gel

### 3. dilution of preparative EcoRV digest for $\sim 20 \text{ ng}/\mu\text{l}$ 1 kb ladder working stock (-NcoI digest):

10 mM Tris-Cl pH 8.0, 0.1 mM EDTA	1.98 ml
EcoRV digest containing $\sim 0.25 \mu\text{g}/\mu\text{l}$ pPSU1 and $\sim 0.25 \mu\text{g}/\mu\text{l}$ pPSU2	0.1 ml
6x gel loading buffer	<u>0.42 ml</u>
	2.5 ml

### 4. dilution of preparative EcoRV digest for $\sim 30 \text{ ng}/\mu\text{l}$ 1 kb ladder working stock (+NcoI digest):

10 mM Tris-Cl pH 8.0, 0.1 mM EDTA	1.93 ml
EcoRV digest containing $\sim 0.25 \mu\text{g}/\mu\text{l}$ pPSU1 and $\sim 0.25 \mu\text{g}/\mu\text{l}$ pPSU2	0.1 ml
NcoI digest containing $\sim 0.25 \mu\text{g}/\mu\text{l}$ pPSU1 and $\sim 0.25 \mu\text{g}/\mu\text{l}$ pPSU2	0.05 ml
6x gel loading buffer	<u>0.42 ml</u>
	2.5 ml

### 5. dilution of preparative PstI digest for $\sim 20 \text{ ng}/\mu\text{l}$ 100 bp ladder working stock:

10 mM Tris-Cl pH 8.0, 0.1 mM EDTA	1.98 ml
PstI digest containing $\sim 0.25 \mu\text{g}/\mu\text{l}$ pPSU1 and $\sim 0.25 \mu\text{g}/\mu\text{l}$ pPSU2	0.1 ml
6x gel loading buffer	<u>0.42 ml</u>
	2.5 ml

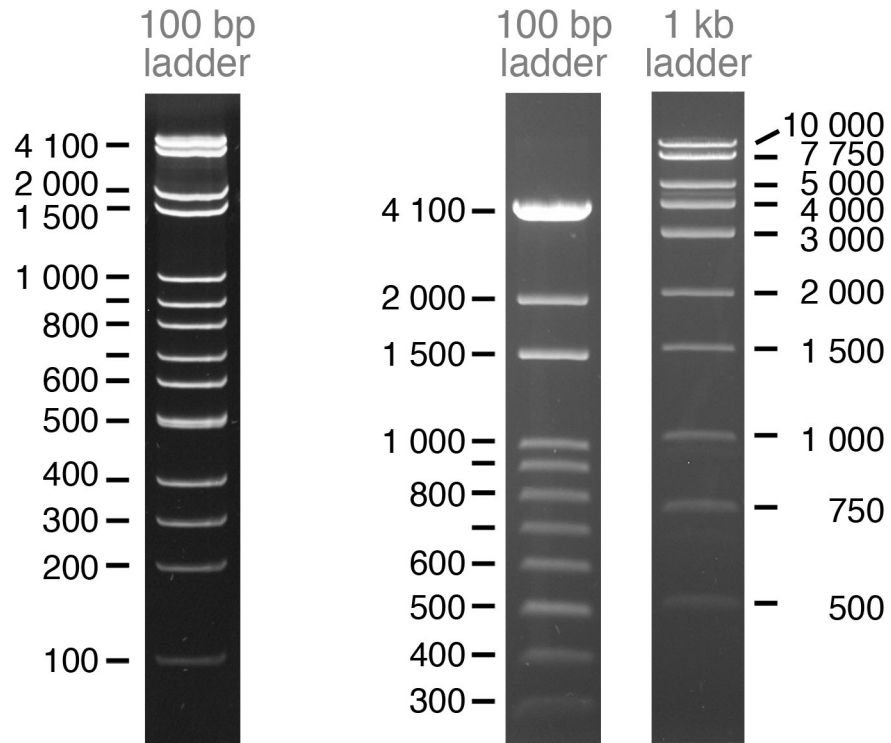
## **Notes**

1. Separate restriction digestions of pPSU1 and pPSU2 are recommended if the concentrations of the DNA concentrations of the two plasmids are not well determined, as might be the case if RNA contamination is present.
2. The amount of restriction enzyme provided is a guide. More or less restriction enzyme may be necessary depending on the quality of the plasmid prep and the source of the restriction enzyme.
3. The suggested working stock of  $\sim 20$  ng/ $\mu$ l is sufficiently concentrated for our needs, but you may wish to use a higher or lower concentration.
4. We use 10-20  $\mu$ l of the 1 kb ladders on agarose gels, and 5  $\mu$ l of the 100 bp ladder on polyacrylamide gels.
5. Gels for the Penn State ladders are provided on p. 3 of this document.

## Penn State DNA ladders

10% acrylamide

1% agarose



The figure above can be printed, cut along the border and inserted into a 5 inch x 7 inch photo holder.